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ATTY.'S DOCKET: ZICHE1

In re Application of:)	Confirmation No. 5690
)	
Marina ZICHE et al)	Art Unit: 1614
)	
Appln. No.: 09/756,185)	Examiner:
)	
Filed: January 9, 2001)	Washington, D.C.
)	
For: COMPOUND B AS ANGIOGENIC...)	November 29, 2004
)	

REQUEST FOR PRIORITY

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Sir:

In accordance with the provisions of 37 CFR §1.55 and the requirements of 35 U.S.C. §119, filed herewith a certified copy of:

EUROPEAN Appln. No.: 98112775.6	Filed: July 9, 1998
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It is respectfully requested that applicant be granted the benefit of the priority date of the foreign application.

Respectfully submitted,

BROWDY AND NEIMARK, P.L.L.C.
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By

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The attached documents are exact copies of the European patent application described on the following page, as originally filed.

Les documents fixés à cette attestation sont conformes à la version initialement déposée de la demande de brevet européen spécifiée à la page suivante.

Patentanmeldung Nr. Patent application No. Demande de brevet n°

98112775.6

Der Präsident des Europäischen Patentamts;
Im Auftrag

For the President of the European Patent Office

Le Président de l'Office européen des brevets
p.o.

R C van Dijk

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Anmeldung Nr.:
Application no.: 98112775.6
Demande no:

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Anmelder/Applicant(s)/Demandeur(s):

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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
If no title is shown please refer to the description.
Si aucun titre n'est indiqué se referer à la description.)

Component B as angiogenic agent in combination with human growth factors.

In Anspruch genommene Priorität(en) / Priority(ies) claimed /Priorité(s)
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COMPOUND B AS ANGIOGENIC AGENT IN COMBINATION WITH HUMAN GROWTH FACTORS

Field of the invention

The present invention refers to the use of Component B as angiogenic agent in combination with human growth factors.

State of the art

Component B (hereinafter indicated as CB) is a 81 amino acid protein originally isolated from human urine. The human gene expressing the protein has been cloned and expressed in CHO cells as recombinant human Component B, the protein has a molecular weight of about 8.9 kD and was thoroughly described in WO 94/14259 to which reference is made also for the methods of preparation and its amino acid sequence.

In WO97/39765 the use of CB as cicatrizant was described.

It is also known that growth factors, as for example basic fibroblast growth factor (bFGF) or vascular endothelial growth factor (VEGF) have angiogenic activity.

Brief description of the drawings

Figure 1: Shows the implant of a pellet loaded with both active compounds (1A) or of two pellets each loaded with one active compound (1B) in the rabbit cornea.

Figure 2: Shows the effect of bFGF and CB on angiogenesis in the rabbit cornea.

Figure 3: Shows the effect of heat-inactivation on the angiogenic activity of CB.

Figure 4: Shows the synergistic effect of CB on bFGF-induced angiogenesis.

Figure 5: Shows the synergistic effect of CB on the angiogenic activity of bFGF.

Figure 6: Represents a drawing of a typical histological section of rabbit cornea illustrating the main structures observable in following figures 7 - 11.

Figure 7: section of rabbit cornea (x200) with a corneal pocket containing 100 ng of bFGF sampled at 6 days post-surgery wherein the arrows show neoformed vessels. (Ep = epithelium).

Figure 8: section of rabbit cornea (x100) with a corneal pocket containing 500 ng of CB sampled at 2 days post-surgery wherein the arrows show neoformed vessels.

Figure 9: section of rabbit cornea (x 200) with a corneal pocket containing 500 ng of CB sampled at 6 days post-surgery wherein the arrows show neoformed

vessels.

Figure 10: Section of rabbit cornea (x100) with a corneal pocket containing 4 µg of CB sampled at 15 days post-surgery wherein the arrow show neoformed vessels.

Figure 11: Section of rabbit cornea (x100) with a corneal pocket containing 500 ng of bFGF sampled at 15 days post-surgery wherein the arrow show neoformed vessels.

Figure 12: mean and 95.0% Tukey HSD Intervals

Detailed description of the invention

It was now surprisingly found that the simultaneous presence of CB and a growth factor increases the angiogenic response elicited by either individual agent, in other words CB is capable of synergizing with a growth factor in promoting neovascular growth in the tissues probably facilitating some early events required to mobilise endothelial cells from capillaries.

Therefore, the main object of the present invention is the use of CB in combination with a growth factor for the manufacture of pharmaceutical compositions useful for the treatment of wounds, ulcers and other traumatic lesions to any of the tissues in the body.

Another object of the invention are pharmaceutical compositions prepared as described above.

A further object of this invention is a method of treatment of wounds, ulcers and other traumatic lesions to any of the tissues in the body, comprising administering an effective amount of CB and of a growth factor, possibly together with a pharmaceutically acceptable excipient.

The administration of the active ingredient may be oral, intravenous, intramuscular, subcutaneous or topical. Other routes of administration, which may establish the desired blood levels of respective active agents are comprised by the present invention.

The administration of the two active compounds can be performed by a single pharmaceutical preparation containing both of them or, preferably, by two pharmaceutical preparation each containing separately one of the two ingredients. Preferred growth factor to be used in combination with CB according to the invention are the basic fibroblast growth factor (bFGF) or the vascular endothelial

growth factor (VEGF).

Angiogenesis was studied in the cornea of albino rabbits since this is an avascular and transparent tissue where inflammatory reactions and growing capillaries can be easily monitored and changes quantified by stereomicroscopic examination (Ziche et al., 1982). This method allows the monitoring over an extended period of time of vessel growth by direct and non traumatic observation of the process. Moreover in the same animal the quantification of the effect can be compared to that of a known agent.

The investigation of the role of Component B (CB) in *in vivo* angiogenesis studied in the rabbit cornea assay was performed by:

- a) testing the ability of the molecule to produce vessel growth when placed into the avascular corneal stroma;
- b) testing the ability of the molecule to favour or repress neovascularization elicited by the angiogenesis factor basic fibroblast growth factor (bFGF) or vascular endothelial growth factor (VEGF).

Methods

Protocol for slow-release preparation of test compounds

Growth factors or peptides were prepared as slow-release pellets.

Slow-release pellets (1x1x0.5 mm) were prepared in sterile conditions incorporating the test substances into a casting solution of a ethinyl-vinyl copolymer (Elvax-40, Dupont, Wilmington, Delaware), in 10% methylene chloride (10 µl/droplet) (Langer and Folkman, 1976; Ziche et al. 1982).

Surgical procedure

The angiogenic activity was assayed *in vivo* using the rabbit cornea assay.

In the lower half of New Zealand white rabbit eye (Charles River, Calco, Lecco, Italy), anaesthetised by sodium pentothal (30 mg/kg), a micro pocket (1.5x3mm) was surgically produced using a pliable iris spatula 1.5 mm wide.

The pellets were implanted in the micropockets located into the transparent avascular corneal stroma.

Quantification of corneal angiogenesis

Subsequent daily observations of the implants were made with a slit lamp stereomicroscope without anaesthesia. An angiogenic response was scored

positive when budding of vessels from the limbal plexus occurred after 3-4 days and capillaries progressed to reach the implanted pellet according to the scheme previously reported (Ziche et al. 1989). Angiogenic activity is expressed as the number of implants exhibiting neovascularization over the total implants studied. Potency is scored by the number of newly formed vessels and by their growth rate. Data are expressed as angiogenesis score, calculated as vessel density x distance from limbus in mm. A density value of 1 corresponded to 0 to 25 vessels per cornea, 2 from 25 to 50, 3 from 50 to 75, 4 from 75 to 100 and 5 for more than 100 vessels (Ziche et al., 1994).

10 Experimental design

The effect of Component B was tested following two procedures:

A) Three different concentrations of the molecule were tested in the cornea of at least 4 distinct rabbits per each dose, to define the potential angiogenic activity of the compound. The effect of Component B was compared with that elicited by the growth factor bFGF at 50 and 100 nag/pellet. In this experimental protocol rabbits were monitored for 3 weeks.

B) To evaluate a potential role of Component B in modulating angiogenesis the effect of this agent was tested in the presence of a defined angiogenesis factor, i.e. bFGF. To this aim two adjacent pockets were surgically produced in the same cornea, one bearing the angiogenic trigger and the other Component B. Experiments were also performed testing both substances incorporated into the same pellet (Figure 1).

This last experimental protocol was specifically set up by our group to define:

1) the effect of the agent as a "costimulator" of the angiogenesis elicited by bFGF; or 2) the ability of the agent to inhibit angiogenesis elicited by the growth factor (Ziche et al, 1992 and 1994).

In this experimental protocol the rabbits were monitored for 4-5 weeks. The same protocol was used to tests the effect of CB as "costimulator" of the angiogenesis induced by VEGF.

30 Histological analysis

Rabbit corneas with corneal pockets containing CB and/or bFGF were sampled at 2, 6, 15 days post surgery, and fixed in formalin after removing the pellets.

Routine histopathological processing was performed; sections 5µm thick were cut next to where each pellet was placed; sections were stained with hematoxylin-eosin. At least 40 sections were examined per each cornea.

Statistical analysis

Results are expressed as means for (n) implants. Angiogenic score data contained both positive and negative results. Multiple comparisons were performed by one-way ANOVA and individual differences were tested by Fisher's test after the demonstration of significant intergroup differences by ANOVA. A P value <0.05 was taken as significant (see also Appendix for further statistical evaluation).

Results

a) Angiogenic activity of CB

The angiogenic activity of CB was tested after incorporating increasing concentrations of the compound in slow release pellets of the polymer Elvax-40. Solubilization and incorporation of the compound into the polymer pellets did not cause any specific problem. The doses tested were: 0.2, 0.5, 2 and 4 µg/pellet. The effect of CB was compared to that produced by basic fibroblast growth factor (bFGF).

CB elicited a dose-dependent angiogenic effect whose potency appeared to be weaker than the one elicited by bFGF. In Figure 2A data are reported on the angiogenic activity of bFGF obtained from previous experiments and from the experiments run in parallel with CB. In Figure 2B data from daily observation of rabbit corneas implanted with CB-containing pellets are reported as angiogenic score. The highest angiogenic score obtained with CB averaged around 3-3.5 (2-4 µg/pellet) (P<0.05 vs vehicle pellets alone) vs 7-8 produced by bFGF (0.2 µg/pellet)(P<0.05 vs vehicle pellets alone). CB was not angiogenic at the concentration of 0.2 µg/pellet. As shown in Table 1, 0.5 µg/pellet CB induced a positive angiogenic response in 1 implant out of 5 performed. Two and 4 µg/pellet were the most effective doses. These doses induced a similar angiogenic activity and produced 2 positive implants out of 5 performed.

CB was devoid of any macroscopic inflammatory activity as revealed by the persistence of corneal transparency all through the experiments at any

concentration tested.

To assess the specificity of CB angiogenic effect, the compound was heat-inactivated (h.i.) by boiling it for 20 min. The dose of 2 μ g was then tested. Following heat inactivation CB completely lost angiogenic activity (Fig. 3)($P < 0.05$ vs CB 2 μ g).

b) Effect of CB on the angiogenesis induced by bFGF

To evaluate the potential role of CB in modulating the effect of a known angiogenic effector, experiments were performed testing suboptimal concentrations of both substances (500 ng of CB and 100 ng of bFGF) co-released into the corneal stroma. Experiments were performed testing both substances incorporated into the same pellet (Fig. 1A). Furthermore the compounds were tested at the same concentration as above but released into the stroma separately in 2 independent pellets (Fig. 1B).

The simultaneous presence of CB and bFGF into the cornea increased the angiogenic response elicited by either individual agent (Fig. 4A and B, Table 2).

Angiogenesis occurred earlier and progressed more rapidly producing a significant increment of the number of newly formed vessels ($P < 0.05$ vs CB and bFGF alone). This effect was apparent in both experimental conditions.

However, when CB and bFGF were released independently by 2 separate pellets the effect was higher. Capillaries grew toward bFGF rather than CB suggesting that CB contributed to potentiate bFGF activity. After 7 days, neovascular growth started to regress.

Additional experiments were performed with increasing concentrations of CB (0.2, 0.5 and 2 μ g/pellet) on the angiogenesis elicited by a constant concentration of bFGF (100 ng). A synergism between the two molecules could be observed (Fig. 5). Interestingly, the most effective condition of synergism between CB and bFGF was observed with 200 ng CB ($P < 0.05$ vs CB and bFGF alone) tested in two separate pellets.

c) Effect of CB on the angiogenesis induced by VEGF

In Table 3 the synergistic effect of Component B on VEGF-induced angiogenesis are reported.

The synergism between CB and VEGF was evaluated with the factors tested in

two separate pellets. The results obtained at day 10 are reported in Table 3. The data are expressed as the number of implants exhibiting neovascularization with an angiogenesis score equal or over to 6, over the total implants performed.

Further statistical analysis was performed in order to confirm possible positive interactions between the test compounds using a more conservative analysis (see Fig. 11 in Appendix).

The main factors ("test compounds" and "angiogenic score" over time) were analysed according to the Multifactor Analysis. The results showed that statistically significant differences ($p < 0.0001$) are present among the test compounds over the time.

As to the interaction among the test compounds the results of the Tukey's test allow the following considerations (see Fig. 12 below):

CB 500 ng + bFGF 100 ng (1 pellet)

Both compounds individually are not statistically different from controls. The combination of the two compounds gave a response which is statistically different from either the controls and the single drugs. The response is around the expected additive effect.

CB 200 ng + bFGF 100 ng (2 pellets)

Both compounds individually are not statistically different from controls. The combination of the two compounds gave a response which is statistically different from either the controls and the single drugs. In addition, it should be noted that the response of the combined treatments clearly exceeds the expected additive effect. The above seems to confirm the presence of a synergistic effect between the two drugs.

CB 500 ng + bFGF 100 ng (2 pellets)

Both compounds are not statistically different from controls. The combination of the two compounds gave a response which is statistically different either from the controls and the single drugs. The response is around the expected additive effect. In addition, no differences were found comparing CB 500 ng + bFGF 100ng (1 pellet) vs CB 500 ng + bFGF 100ng (2 pellet).

CB 2 μ g + bFGF 100 ng (2 pellets)

Both compounds are not statistically different from controls. The combination of

the two compounds gave a response which is statistically different from controls but not from the single compounds. The response is around the expected additive effect.

Histological analysis

5 The effect of CB was examined at the maximal effective concentration (4 μ g) and at a suboptimal concentration (500 ng) in the presence and in the absence of bFGF (100 ng). No difference in the extent of cellular infiltrate was apparent between CB and bFGF implants in any combination (Figs. 7,8,9). Within 2 days from the implants a leukocyte infiltrate surrounded a dense network of newly
10 formed capillaries in proximity of the limbal region at the epithelial side of the cornea (Fig. 10). At day 6 a consistent reduction in the extent of the leukocyte infiltrate was apparent while capillary vessels appeared increased in number and caliber in response to either molecule (Fig. 11). At day 15 the extent of the leukocyte infiltrate was negligible while capillaries appeared morphologically
15 unmodified.

Conclusions

Component B possesses angiogenic activity which is apparent in the concentration range of micrograms and which is lost by heat inactivation. Most of the angiogenesis factors are angiogenic at concentrations 20-40 fold lower.
20 Together with the high concentration required to elicit angiogenesis, 2 aspects appear relevant in CB effect:

- 1) the ability to elicit budding of capillaries within the first 3-4 days from the implant, mimicking the secreted angiogenesis factor VEGF rather than the matrix linked angiogenesis factor bFGF;
- 25 2) the flattening over time of the efficiency of neovascular growth, leading to only 30-40% of the implant tested to be fully vascularized after 10-14 days.

Our results indicate that CB induces angiogenesis *in vivo* and has the ability to synergize with bFGF in promoting neovascular growth in the rabbit cornea.

These considerations together with the characteristics of the potentiation of the
30 angiogenic response in the presence of bFGF, suggest that CB requires the presence of additional growth factors to fully express its angiogenic potential *in vivo*.

Histological examination of corneal sections sampled at various time intervals was performed to assess whether the angiogenesis process elicited by CB involved inflammatory cell infiltrate. The effect of CB was compared to that produced by the corneal implant of bFGF. At routine histological examination we did not find major differences in the extent and in the type of leukocyte infiltrate in corneas receiving CB, bFGF or the combination of the two. Thus from our results we can conclude that the corneal vascularization induced by CB does not appear to be mediated by gross inflammatory reaction products since no sign of corneal opacity was apparent.

- 10 The characteristic of the angiogenic response elicited in the avascular cornea by CB suggests that CB might facilitate some of the early events required to mobilise endothelial cells from capillaries. Once this process is started and endothelial cells are "loosened" from the tight boundary to the extracellular matrix, bFGF expresses its mitogenic effect with more efficiency.

Table 1 Effect of Component B on angiogenesis in the rabbit cornea

CB (µg/pellet)	Positive implants/total performed	bFGF (ng/pellet)	Positive implants/total performed
0	0/6	0	0/6
0.2	0/5	50	1/6
0.5	1/5	100	2/6
2	2/5	200	5/6
4	2/5		

Data are expressed as positive implants exhibiting neovascularization over the total implants performed. The results obtained at day 7 are reported.

An angiogenic response was scored positive when budding of vessels from the limbal plexus occurred after 3-4 days and capillaries progressed to reach the pellet containing the angiogenic factors.

Table 2 Synergistic effect of Component B on bFGF-induced angiogenesis

Test compounds	Positive implants/total performed
CB 500 ng	1/5
bFGF 100 ng	1/5
CB 500 ng + bFGF 100 ng (one pellet)	3/6
CB 500 ng + bFGF 100 ng (two pellets)	4/5

Data are expressed as positive implants exhibiting neovascularization over the total implants performed. The results obtained at day 7 are reported.

An angiogenic response was scored positive when budding of vessels from the limbal plexus occurred after 3-4 days and capillaries progressed to reach the pellet containing the angiogenic factors.

TABLE 3

Synergistic effect of Component B on VEGF-induced angiogenesis

Positive implants/total performed	
CB 200 ng	0/5
CB 500 ng	1/5
VEGF 100 ng	1/4
CB 200 ng + VEGF 100 ng	2/4
CB 400 ng + VEGF 100 ng	4/4

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CLAIMS

1. Use of Component B in combination with a human growth factor for the preparation of a composition for promoting angiogenesis.
2. The use according to Claim 1 in the treatment of wounds, ulcers and other
5 traumatic lesions to any of the tissues in the body.
3. Pharmaceutical composition useful as cicatrizants comprising Component B and a human growth factor as active principles in combination with a pharmaceutically acceptable carrier.
4. Pharmaceutical composition according to Claim 3 wherein the two active
10 principle are both present in a single administration dose.
5. Pharmaceutical composition according to Claim 3 wherein the two active principle are present each in a separated administration dose.
6. Pharmaceutical composition according to Claims 3 - 5 wherein the human growth factor is bFGF or VEGF.
- 15 7. Method of treatment of wounds, ulcers and other traumatic lesions to any of the tissues in the body comprising administering in a single administration dose an effective amount of Component B and an effective amount of a human growth factor.
8. Method according to Claim 7 wherein the two active principles are administered
20 in separated administration dose.
9. Method according to Claims 8 and 9 wherein the human growth factor is bFGF or VEGF.

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COMPOUND B AS ANGIOGENIC AGENT IN COMBINATION WITH HUMAN
GROWTH FACTORS

Abstract

The present invention refers to the use of Compound B as angiogenic agent in
5 combination with human growth factors.

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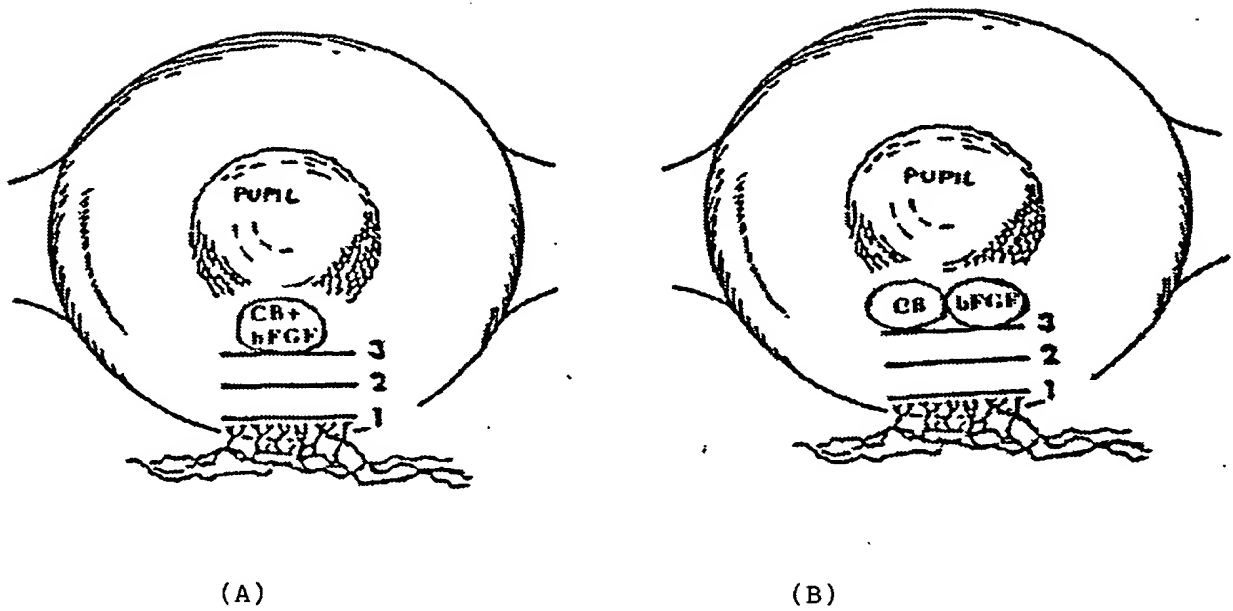


Figure 1

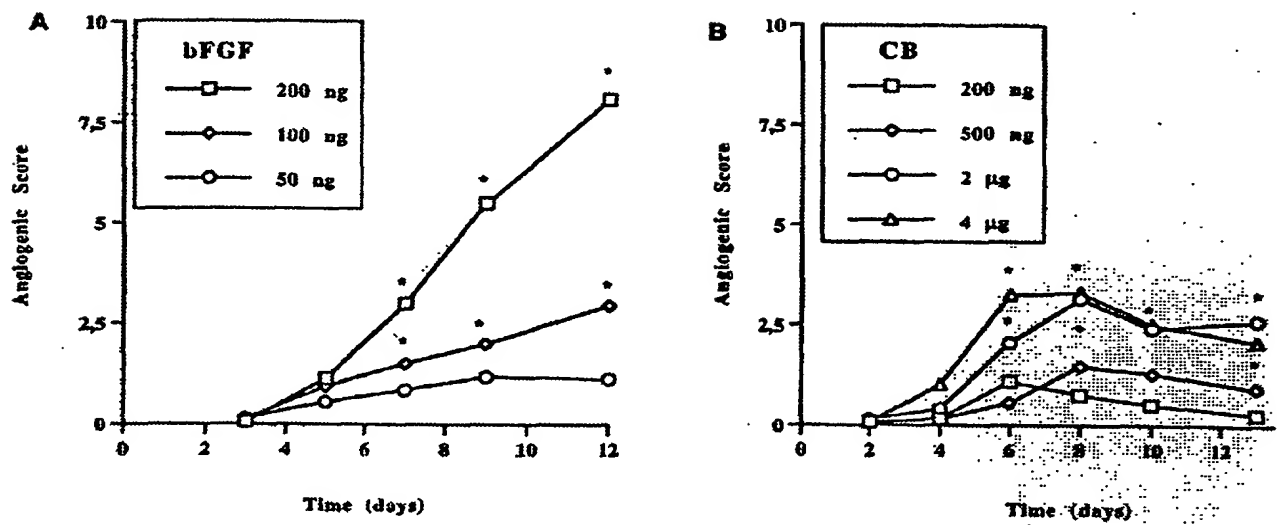


Figure 2

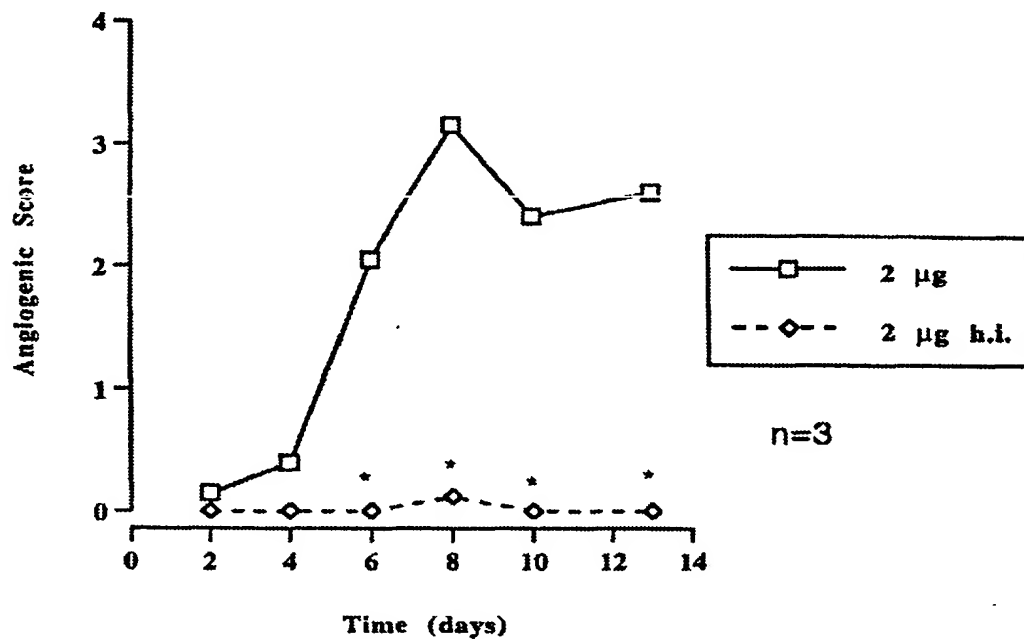


Figure 3

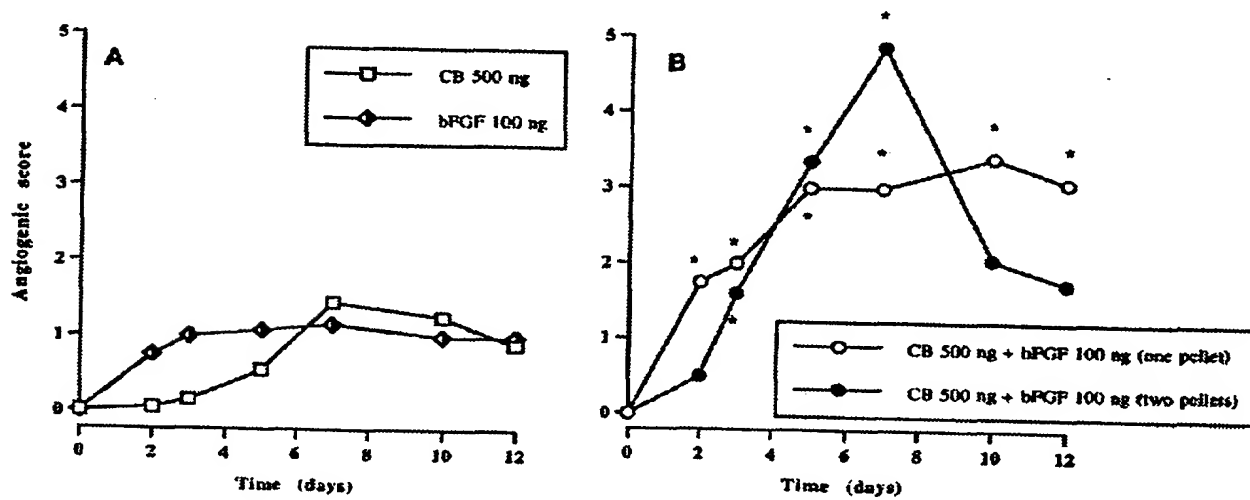


Figure 4

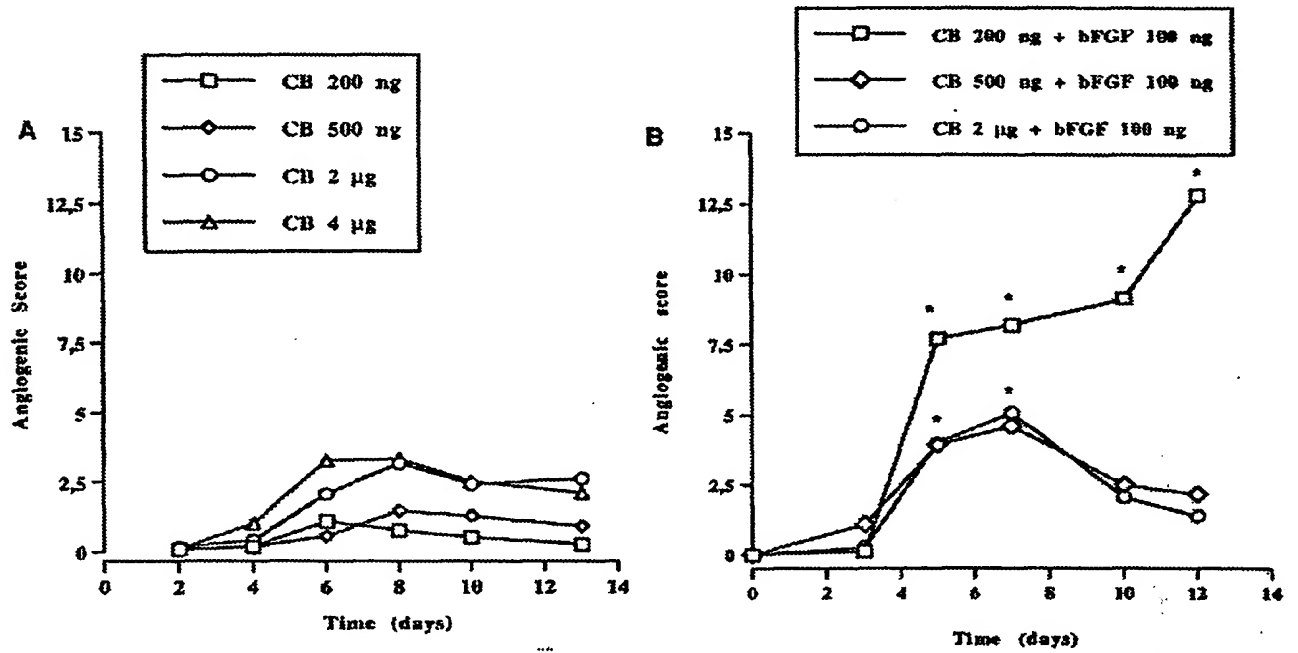


Figure 5

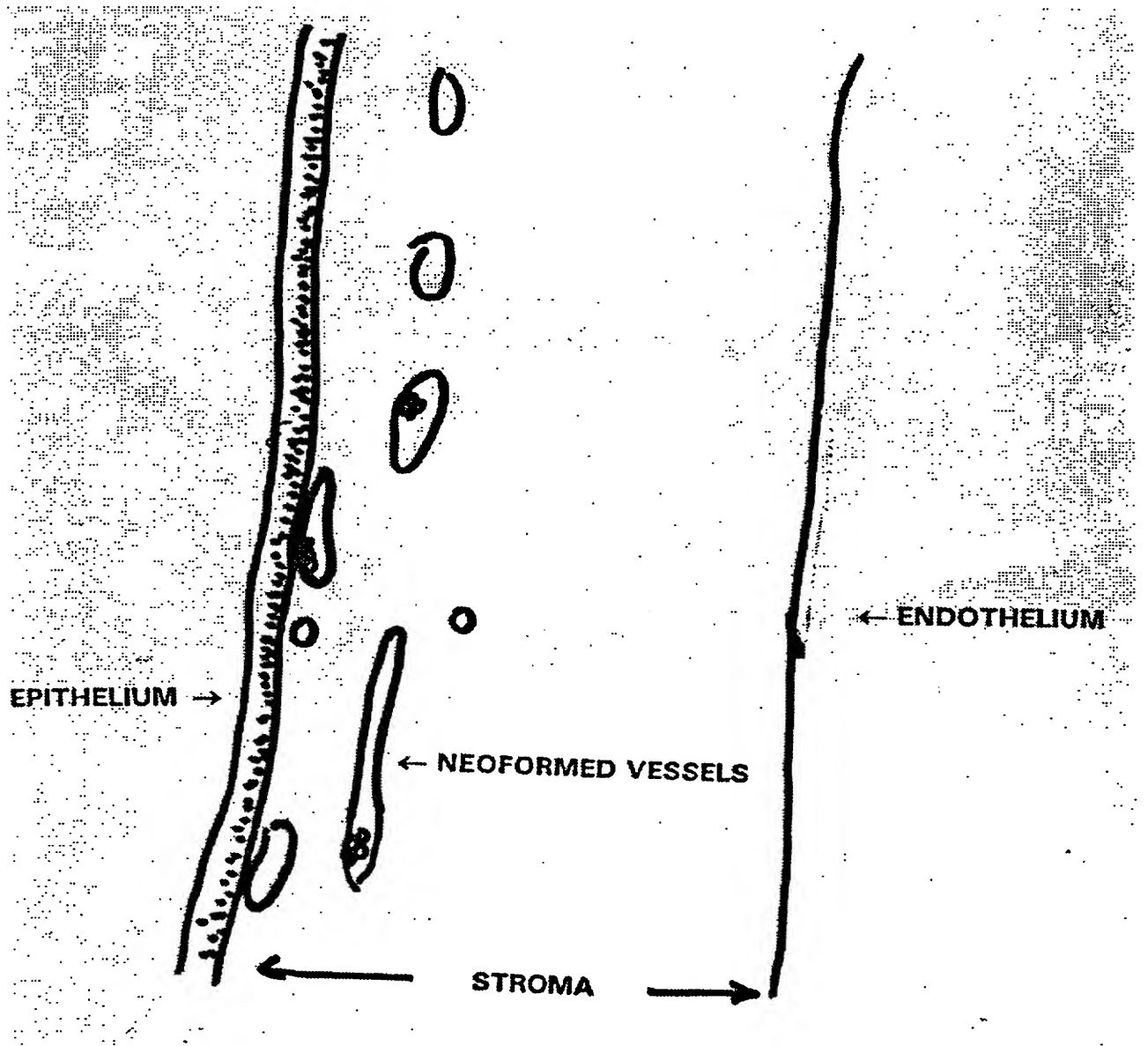


Figure 6

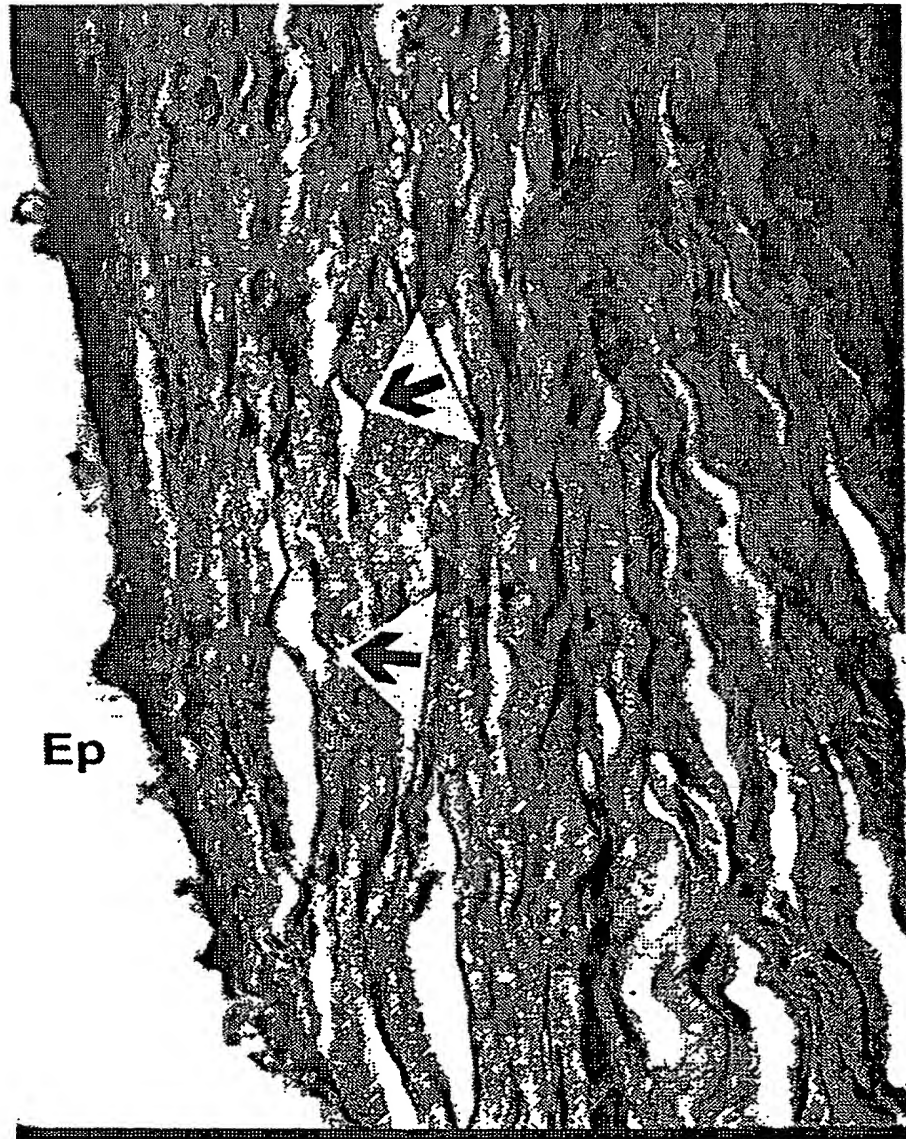


Figure 7

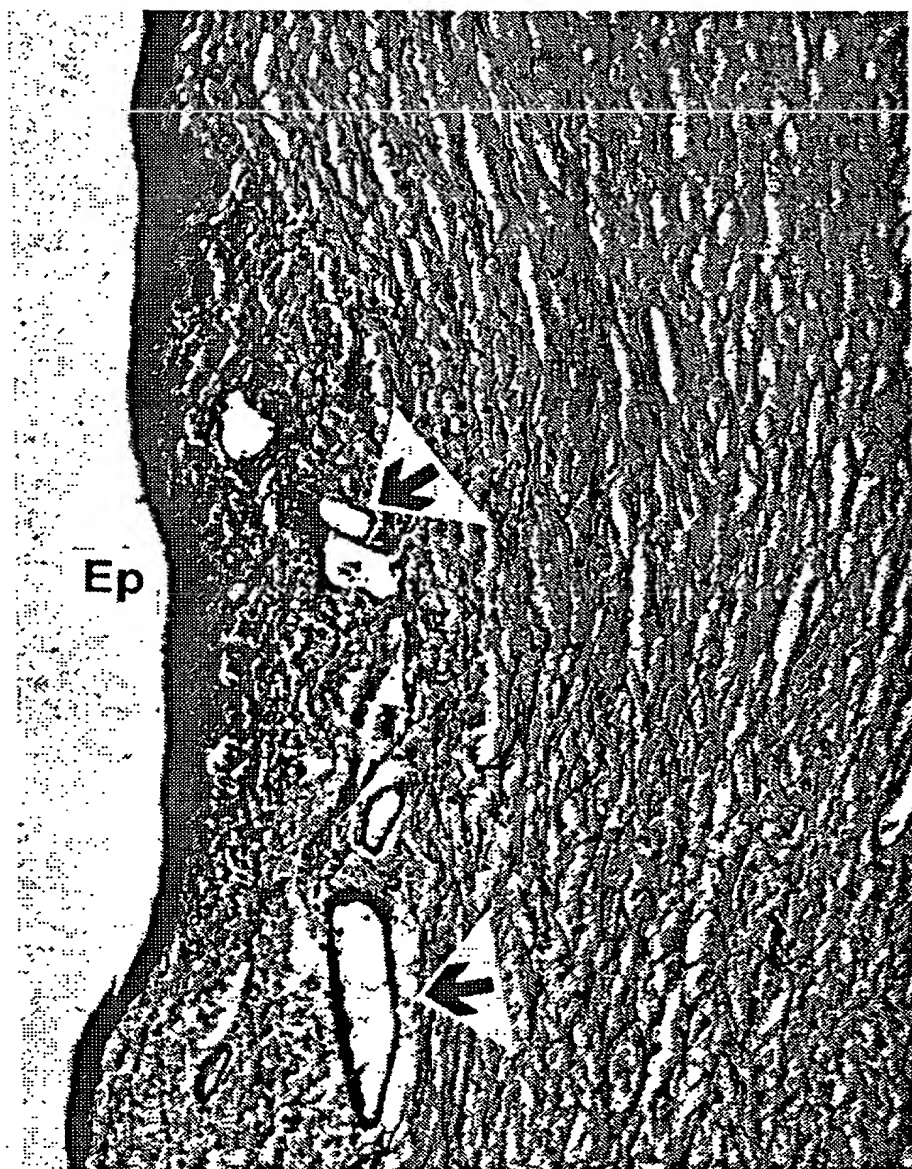


Figure 8

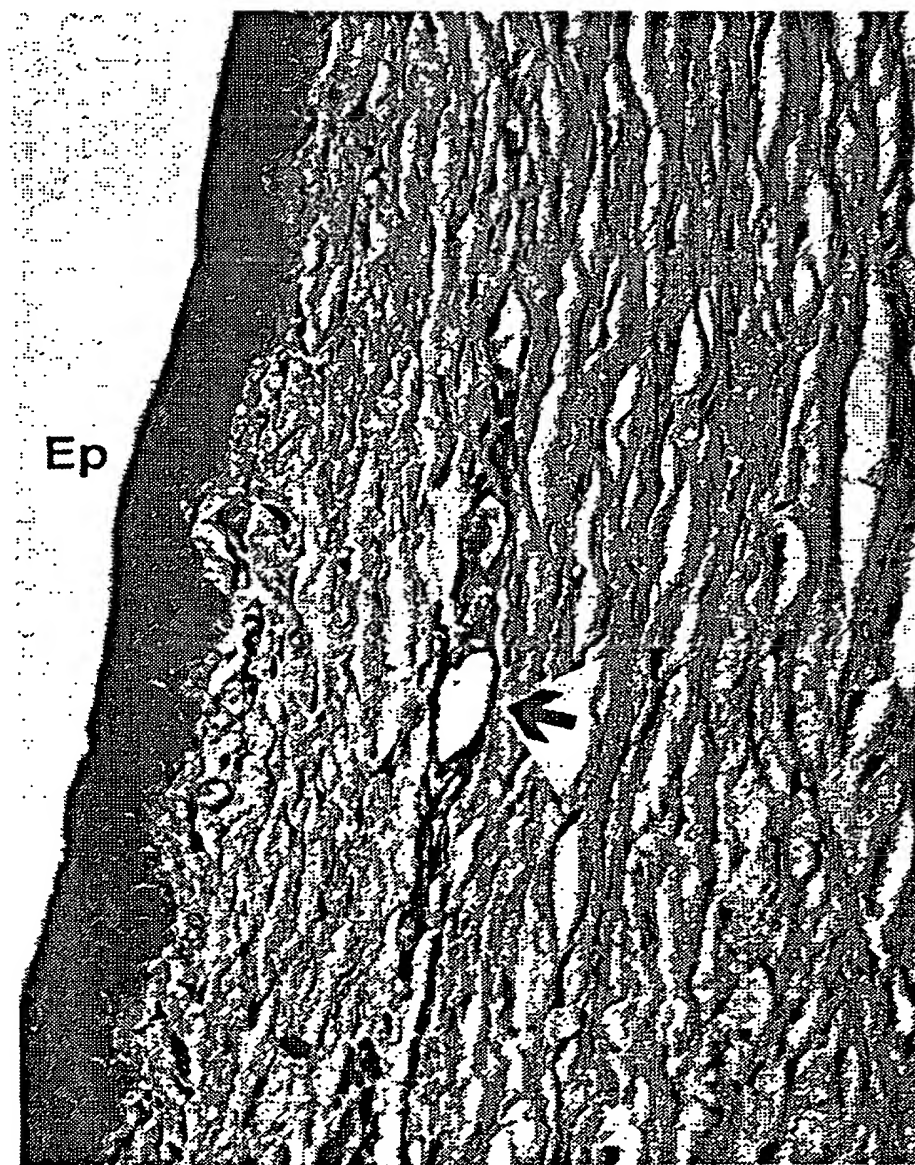


Figure 9

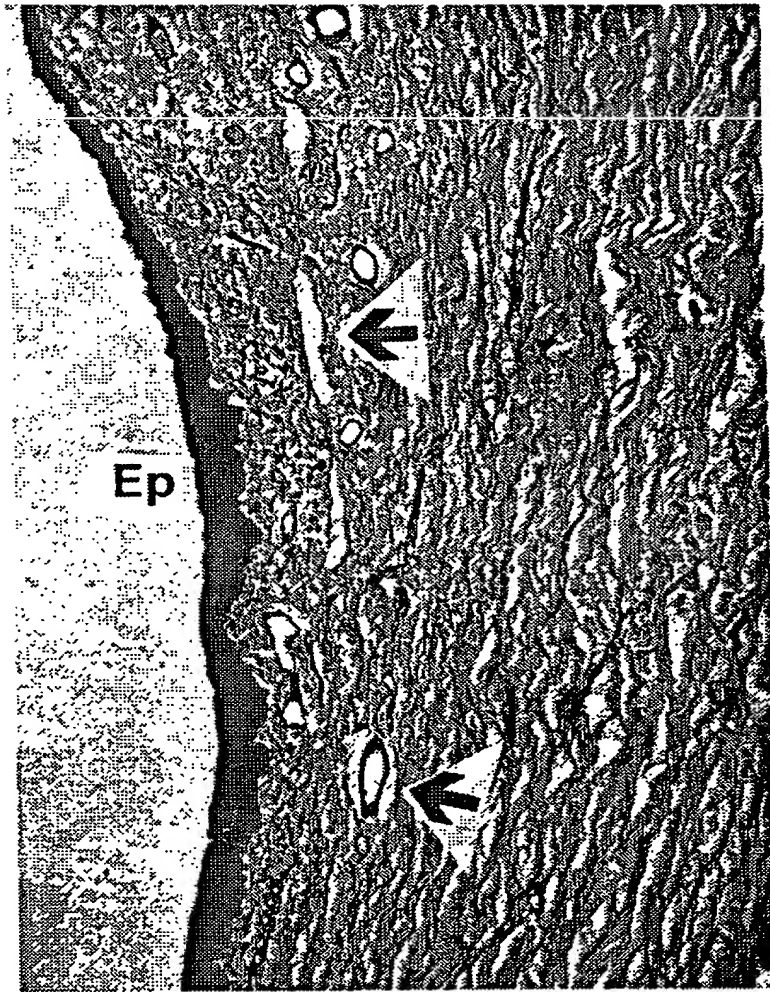


Figure 10

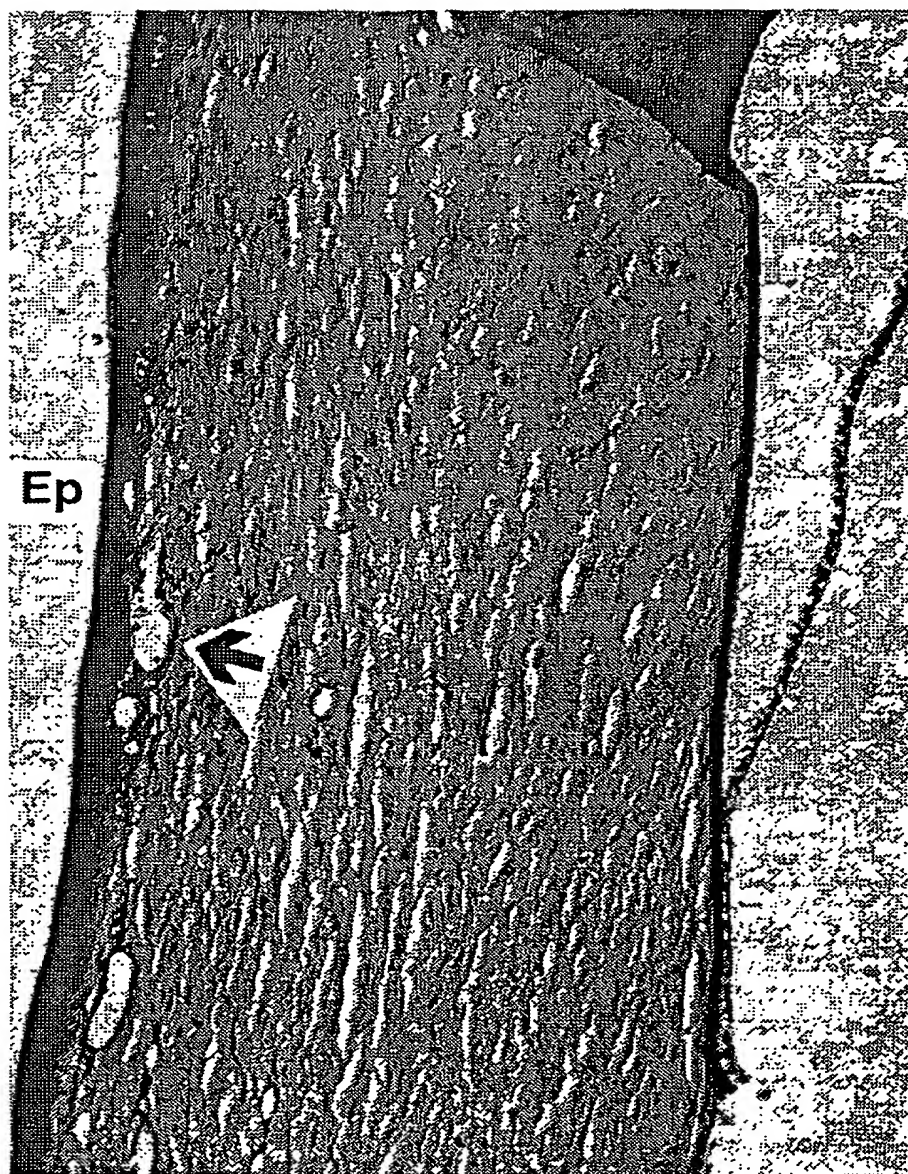


Figure 11

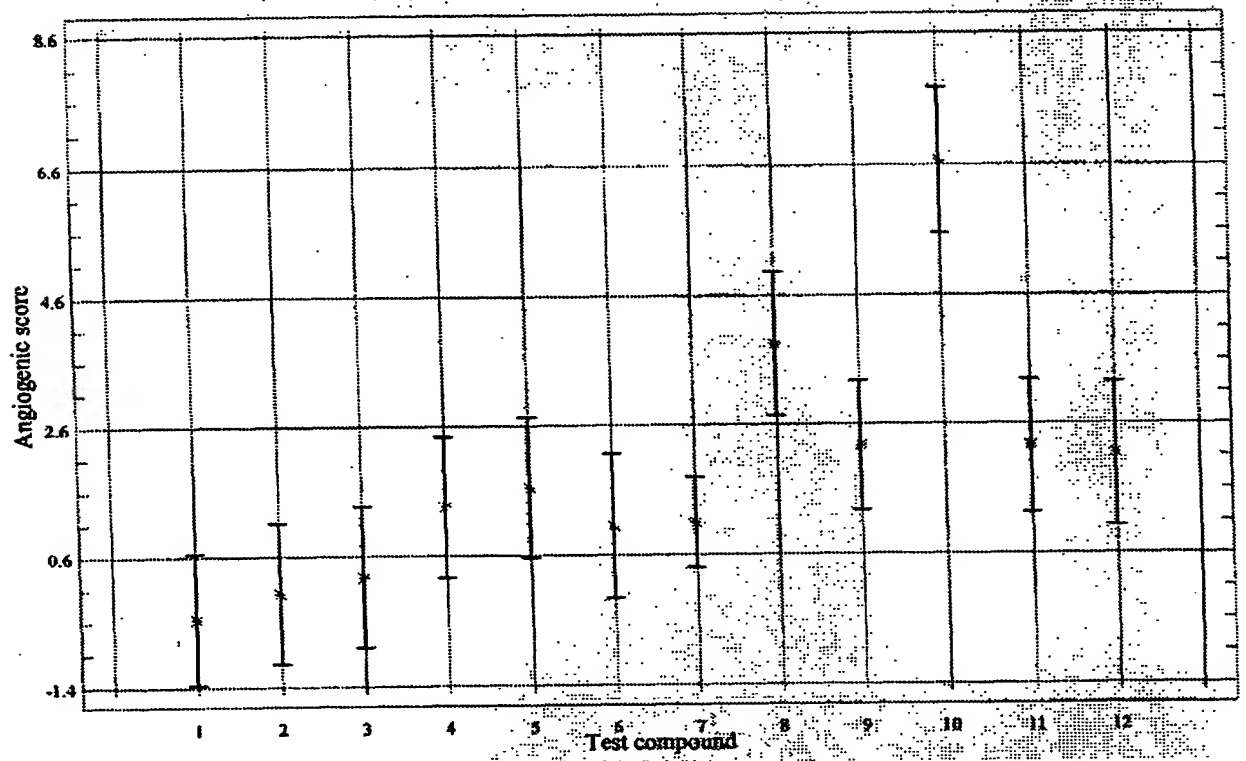


Figure 12

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